



合肥万物生物科技有限公司  
HEFEI WANWU technology CO., LTD

### 冰冻切片荧光 tunel 实验报告

#### 1. 实验器材及试剂

##### 1.1 实验器材

| 名称      | 厂家           | 型号                  |
|---------|--------------|---------------------|
| 冰冻切片机   | Thermo       | Cryotome E          |
| 载玻片     | wanwu        |                     |
| 盖玻片     | 江苏世泰实验器材有限公司 | 10212432C           |
| 脱色摇床    | wanwu        | TSY-B               |
| 涡旋混合器   | wanwu        | MX-F                |
| 掌上离心机   | wanwu        | D1008E              |
| 移液枪     | Dragon       | KE0003087/KA0056573 |
| 组化笔     | wanwu        | WG1066-1            |
| 冰箱      | 青岛海尔股份有限公司   | BCD-192TGN          |
| 正置荧光显微镜 | 日本尼康         | NIKON ECLIPSE C1    |
| 成像系统    | 日本尼康         | NIKON DS-U3         |

##### 1.2 主要实验试剂

| 试剂        | 厂家    | 货号    |
|-----------|-------|-------|
| 4%多聚甲醛    | wanwu | G1101 |
| PBS 缓冲液   | wanwu | G0002 |
| 蛋白酶 K     | wanwu | G1205 |
| 破膜液       | wanwu | G1204 |
| DAPI      | wanwu | G1012 |
| 抗荧光淬灭封片剂  | wanwu | G1401 |
| Tunel 试剂盒 | wanwu | G1501 |



## 2.冰冻切片荧光 tunel 实验步骤

- 2.1 冰冻切片固定：冰冻切片 37°C 烘箱烘烤 10-20min，控干水分。置于 4% 多聚甲醛固定 30min，于 PBS (PH7.4) 中在脱色摇床上晃动洗涤 3 次，每次 5min。
- 2.2 蛋白酶 K 修复：切片稍甩干后用组化笔在组织周围画圈（防止液体流走），在圈内滴加蛋白酶 K 工作液覆盖组织，37 度温箱孵育 22min。将玻片置于 PBS (PH7.4) 中在脱色摇床上晃动洗涤 3 次，每次 5min。（蛋白酶 K 工作液配置方法，原液：PBS=1:9）
- 2.3 破膜：切片稍甩干后在圈内滴加破膜工作液覆盖组织，常温下孵育 20min，将玻片置于 PBS (PH7.4) 中在脱色摇床上晃动洗涤 3 次，每次 5min。（破膜液为 0.1% triton. 配置方法，triton 原液：PBS=1:1000）
- 2.4 室温平衡：切片稍甩干后在圈内滴加 buffer 覆盖组织，buffer 常温孵育 10min.
- 2.5 加反应液：按片子数量和组织大小取 tunel 试剂盒内适量 TDT 酶 ,dUTP, buffer 按 1:5:50 比例混合，加到圈内覆盖组织，切片平放于湿盒内，37°C 恒温箱孵育 2 小时，湿盒内加少量水保持湿度。
- 2.6 DAPI 复染细胞核：切片用 PBS (PH7.4) 洗涤 3 次，每次 5min。去除 PBS 后在圈内滴加 DAPI 染液，避光室温孵育 10min。
- 2.7 封片：玻片置于 PBS (PH7.4) 中在脱色摇床上晃动洗涤 3 次，每次 5min。切片稍甩干后用抗荧光淬灭封片剂封片。
- 2.8 镜检拍照：切片于荧光显微镜下观察并采集图像。（DAPI 紫外激发波长 330-380nm，发射波长 420nm，发蓝光；FITC 激发波长 465-495nm，发射波长 515-555 nm，发绿光；）

## 3.冰冻切片荧光 tunel 结果判读

DAPI 染出来的细胞核在紫外的激发下为蓝色，试剂盒为 FITC 荧光素标记，阳性凋亡细胞核为绿色



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### (FITC) tunel assay protocol (frozen-slides)

#### 1. Apparatus and reagents

##### 1.1 Apparatus

| Name                    | Producer | Model               |
|-------------------------|----------|---------------------|
| Freezing microtome      | Thermo   | Cryotome E          |
| Glass microscope slides | wanwu    |                     |
| Coverslips              | CITOTEST | 10212432C           |
| Rocker                  | wanwu    | TSY-B               |
| Vortex                  | wanwu    | MX-F                |
| Micro-centrifuge        | wanwu    | D1008E              |
| Pipettor                | Dragon   | KE0003087/KA0056573 |
| Liquid blocker pen      | wanwu    | WG1066-1            |
| Refrigerator            | Haier    | BCD-192TGN          |
| Ortho-Fluorescent       | Nikon    | NIKON ECLIPSE C1    |
| Microscopy              |          |                     |
| Imaging system          | Nikon    | NIKON DS-U3         |

##### 1.2 Major reagents

| Name                      | Producer | Model |
|---------------------------|----------|-------|
| 4% paraformaldehyde       | wanwu    | G1101 |
| PBS solution              | wanwu    | G0002 |
| Proteinase K              | wanwu    | G1205 |
| Permeabilize solution     | wanwu    | G1204 |
| DAPI                      | wanwu    | G1012 |
| anti-fade mounting medium | wanwu    | G1401 |
| Tunel assay kit           | wanwu    | G1501 |

网址 : <http://www.hfwanwu.com>

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地址 : 合肥市蜀山区长江西路248号11层

电话 : 400-1016-218

邮箱 : 2028438226@qq.com



## 2. Procedure

2.1 Fix frozen-slides: restore frozen-slides to room temperature and dry in air. Fix in cold acetone 10 min and then dry in air. Wash three times with PBS (pH 7.4) in a Rocker device, 5 min each

2.2 Antigen retrieval: eliminate obvious liquid, mark the objective tissue with liquid blocker pen. Add proteinase K working solution to cover objectives and incubate at 37°C for 25 min. then wash three times with PBS (pH 7.4) in a Rocker device, 5 min each. (Method for configuring working solution of proteinase K, stock solution: PBS=1:9)

2.3 Permeabilization: eliminate excess liquid, add permeabilize working solution to cover objective tissue, then incubate at room temperature for 20 min. wash three times with PBS (pH 7.4) in a Rocker device, 5 min each. (The membrane breaking fluid is 0.1% triton. Configuration method, triron stock solution: PBS=1:1000)

2.4 Equilibrium at room temperature: After the slices are slightly dried, buffer is added to the tissues in the circle, and the buffer is incubated at room temperature for 10 minutes

2.5 Tunel reaction: Take appropriate amount of TDT enzyme, dUTP and buffer in the tunel kit according to the number of slices and tissue size and mix at 1:5:50 ratio。 Prepare this reaction solution according to demand before use. Add this mixture to objective tissue placed in a flat wet box, incubate at 37°C for 2 h. be sure to keep the wet box moist by adding water.

2.6 DAPI counterstain in nucleus: wash three times with PBS (pH 7.4) in a Rocker device, 5 min each. Then incubate with DAPI solution at room temperature for 10 min, kept in dark place

2.7 Mount: wash three times with PBS (pH 7.4) in a Rocker device, 5 min each. Throw away liquid slightly, then coverslip with anti-fade mounting medium.

2.8 Microscopic examination and collecting images through fluorescence microscope. DAPI emits blue light at an ultraviolet excitation wavelength of 330-380 nm and an emission wavelength of 420 nm; FITC has an excitation wavelength of 465-495 nm and an emission wavelength of 515-555 nm, and emits green.

## 3. Results

Nucleus is blue by labeling with DAPI. Tunel assay kit is from Roch and labelled with FITC. Positive apoptosis cells are green.